

Doktoranto Karolio Šablauskio ataskaita už
2024/2025 mokslo metų pirmą pusmetį

- **Disertacijos pavadinimas:** Genetinių pokyčių charakterizavimas naudojant giliojo mokymo neuroninius tinklus (angl. Characterization of genetic changes using deep neural networks)
- **Darbo vadovas:** prof. Audronė Jakaitienė
- **Doktorantūros pradžios ir pabaigos metai:** 2022 – 2027 (akademinės atostogos 2024-10-01 – 2025-01-31)
- **Studijų metai:** 3.

Studijų metai	Egzaminai	
	Planas	Įvykdyta
I (2022/2023)	1	1
II (2023/2024)	2	2
<i>Akademinės atostogos</i> 2024-10-01 – 2025-01-31		
III (2025/2026)	1	0
IV (2026/2027)	0	0
Iš viso:	4	3

Išlaikytas egzaminas dalykui "Didžiųjų duomenų analitika".

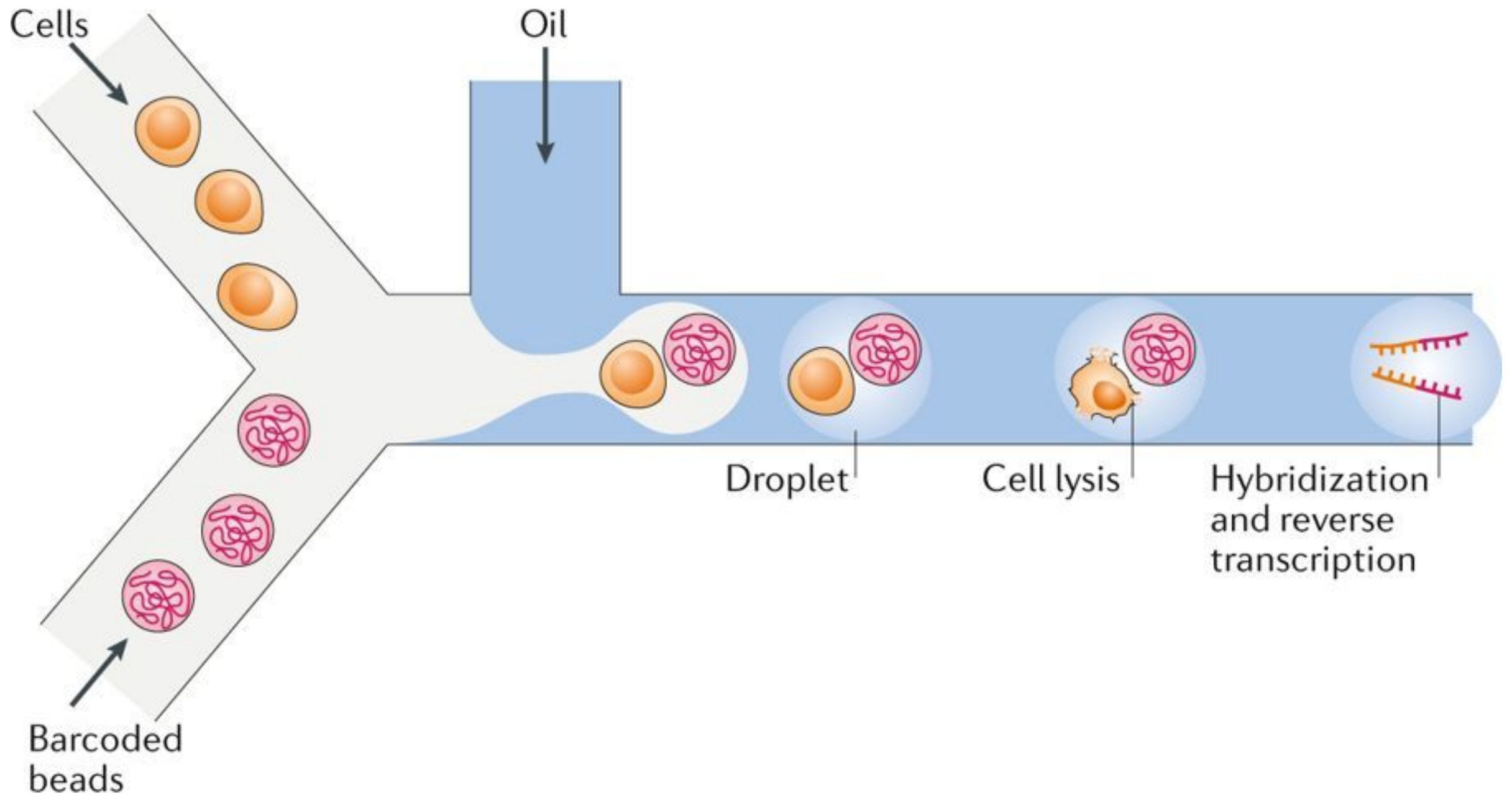
Studijų metai	Dalyvavimas konferencijose				Publikacijos					
	Tarptautinėse		Nacionalinėse		Su citavimo rodikliu			Be citavimo rodiklio		
	Planas	Ivykdyta	Planas	Ivykdyta	Planas	Ivykdyta	Būklė	Planas	Ivykdyta	Būklė
I (2022/2023)	0	0	0	0	0	0	0	0	0	0
II (2023/2024)	1	0	0	0	0	0	0	0	0	0
Akademines atostogos 2024-10-01 – 2025-01-31										
III (2025/2026)	1	0	0	0	1	0	1 pateikta publikacija	0	0	-
IV (2026/2027)	1	0	0	0	1	0		0	0	-
Iš viso:	3	0	0	0	2	0		0	0	-

1 High-throughput single cell -omics using semi-permeable capsules
2
3 Denis Baronas¹, Justina Zvirblyte¹, Simonas Norvaisis¹, Greta Leonaviciene¹, Karolis Goda¹,
4 Vincenta Mikulenaite¹, Vytautas Kaseta³, Karolis Sablauskas^{2,4}, Laimonas Griskevicius²,
5 Simonas Juzenas¹ and Linas Mazutis^{1,5,*}
6
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8 ² Hematology, Oncology and Transfusion Medicine Center, National Cancer Center, Vilnius University Hospital
9 Santaros Clinics, Vilnius, Lithuania
10 ³ Institute of Data Science and Digital Technologies, Vilnius University, Vilnius, Lithuania
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12 ⁵ Department of Molecular Biology, Umea University, Sweden
13 * Correspondence
14

15 **Abstract**

16
17 Biological systems are inherently complex and heterogeneous. Deciphering this
18 complexity increasingly relies on high-throughput analytical methods and tools that efficiently
19 probe the cellular phenotype and genotype. While recent advancements have enabled various
20 single-cell -omics assays, their broader applications are inherently limited by the challenge of
21 efficiently conducting multi-step biochemical assays while retaining various biological
22 analytes. Extending on our previous work (*1*) here we present a versatile technology based on
23 semi-permeable capsules (SPCs), tailored for a variety of high-throughput nucleic acid assays,
24 including digital PCR, genome sequencing, single-cell RNA-sequencing (scRNA-Seq) and
25 FACS-based isolation of individual transcriptomes based on nucleic acid marker of interest.
26 Being biocompatible, the SPCs support single-cell cultivation and clonal expansion over long
27 periods of time – a fundamental limitation of droplet microfluidics systems. Using SPCs we
28 perform scRNA-Seq on white blood cells from patients with hematopoietic disorders and
29 demonstrate that capsule-based sequencing approach (CapSeq) offers superior transcript
30 capture, even for the most challenging cell types. By applying CapSeq on acute myeloid
31 leukemia (AML) samples, we uncover notable changes in transcriptomes of mature
32 granulocytes and monocytes associated with blast and progenitor cell phenotypes. Accurate
33 representation of the entirety of the cellular heterogeneity of clinical samples, driving new
34 insights into the malfunctioning of the innate immune system, and ability to clonally expand
35 individual cells over long periods of time, positions SPC technology as customizable, highly
36 sensitive and broadly applicable tool for easy-to-use, scalable single-cell -omics applications.
37
38

- Išlaikytas 1 egzaminas (3/4) - dar 1 numatomas šį semestrą
- Pateiktas manuskriptas publikacijai su GMC tyrėjais žurnalui *Nature* (IF 50.5). Autorių sąrašė 8/11.
- Ruošiama publikacija pirmuoju autorium į *Bioinformatics* (IF 6.9) arba *Nucleic Acid Research* (16.7).
- Konferencijos - 1 abstraktas buvo pateiktas, bet nebuvo priimtas. Numatoma teikti 2 šį semestrą.
- Stažuotė numatoma 2025-05 mėn.



<https://www.rna-seqblog.com/single-cell-rna-sequencing-for-the-study-of-development-physiology-and-disease/>

DATA STRUCTURE

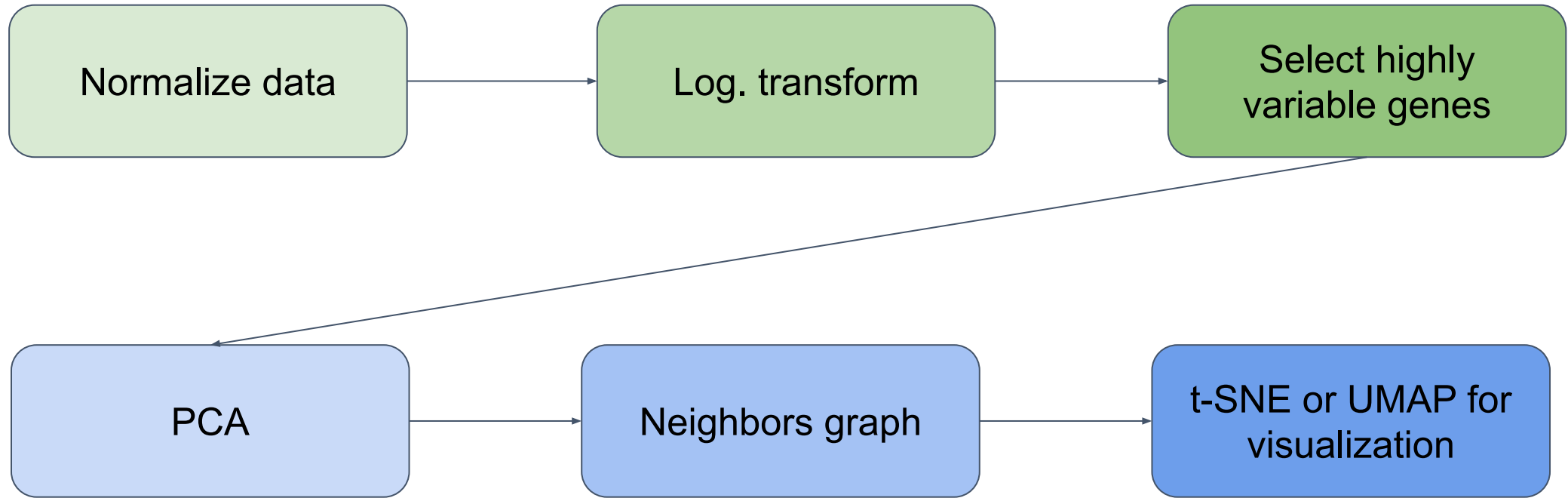
	cell_0	cell_1	...	cell_N
gene_0	12	0		180
gene_1	0	20		0
...				
gene_M	0	15		0

matrix **M x N** - **Count matrix**

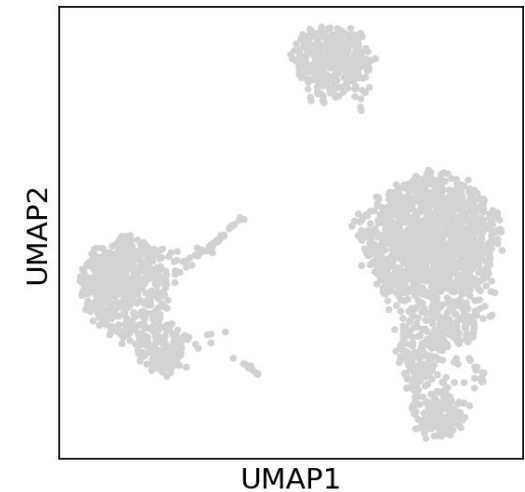
$M = 2 \cdot 10^3 - 3 \cdot 10^4$

$N = 2 \cdot 10^3 - 2 \cdot 10^6$

COUNT MATRIX PROCESSING



DIMENSIONALITY REDUCTION



Challenges related to current approach:

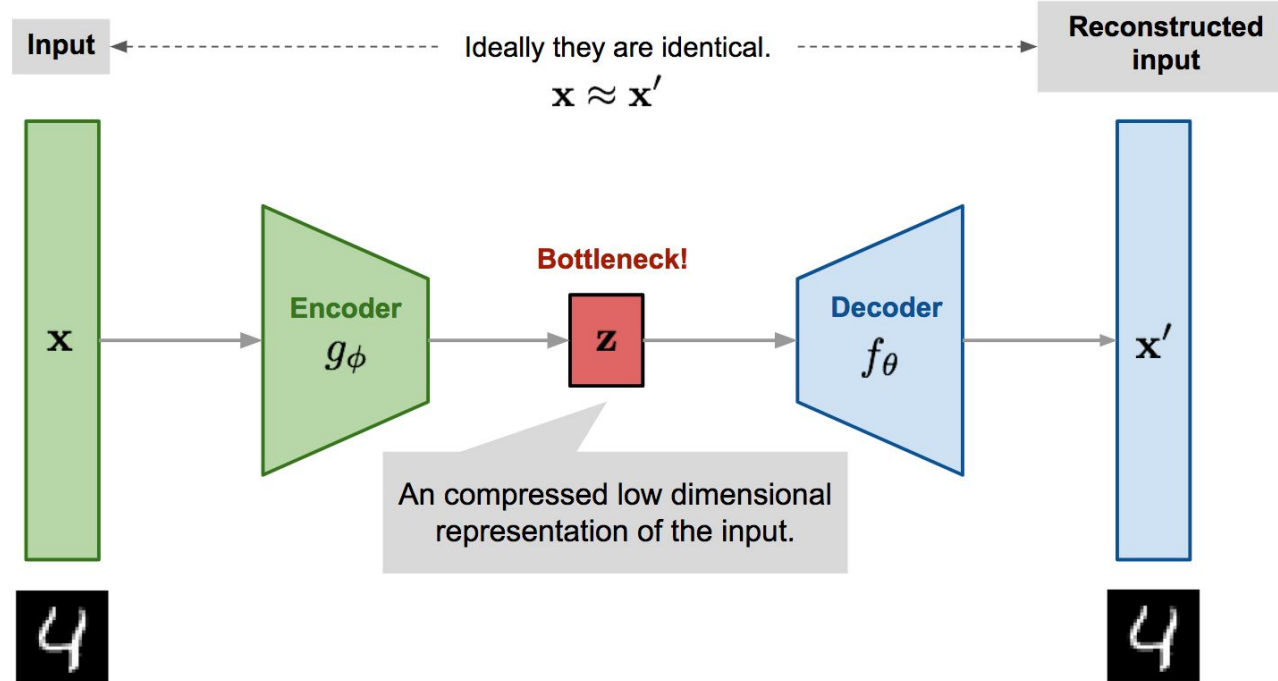
- a number of parameters are chosen in an **arbitrary** way
- each change requires **regenerating** projection

Step	Change in the number of dimensions	Reasoning	Is the choice of parameters arbitrary?
Selection of variable genes - biological priors	37,773 → 37,528	Mitochondrial and ribosomal genes are non-differentiating so these features can be removed	No - mitochondrial and ribosomal gene sets are well defined.
Selection of variable genes - metric-based approach	37,528 → 3,000	Fano factor (or other variance metric) allows selecting highly variable or highly informative genes	Depends on the specific approach used. In the case of the Fano factor - yes.
PCA	3,000 → 93	Selects only the highly informative principal components	No - number of principal components can be chosen using the elbow method
KNN	not applicable	Constructs a graph of k nearest neighbors for each cell.	Yes - “rules of thumb” exist, for example k should be the minimum number of cells expected for each type.
UMAP	93 → 2	Computes the local and global structure of the data.	Yes - requires experimentation to find optimal spacing of cells.

2000 dimensions

2 dimensions

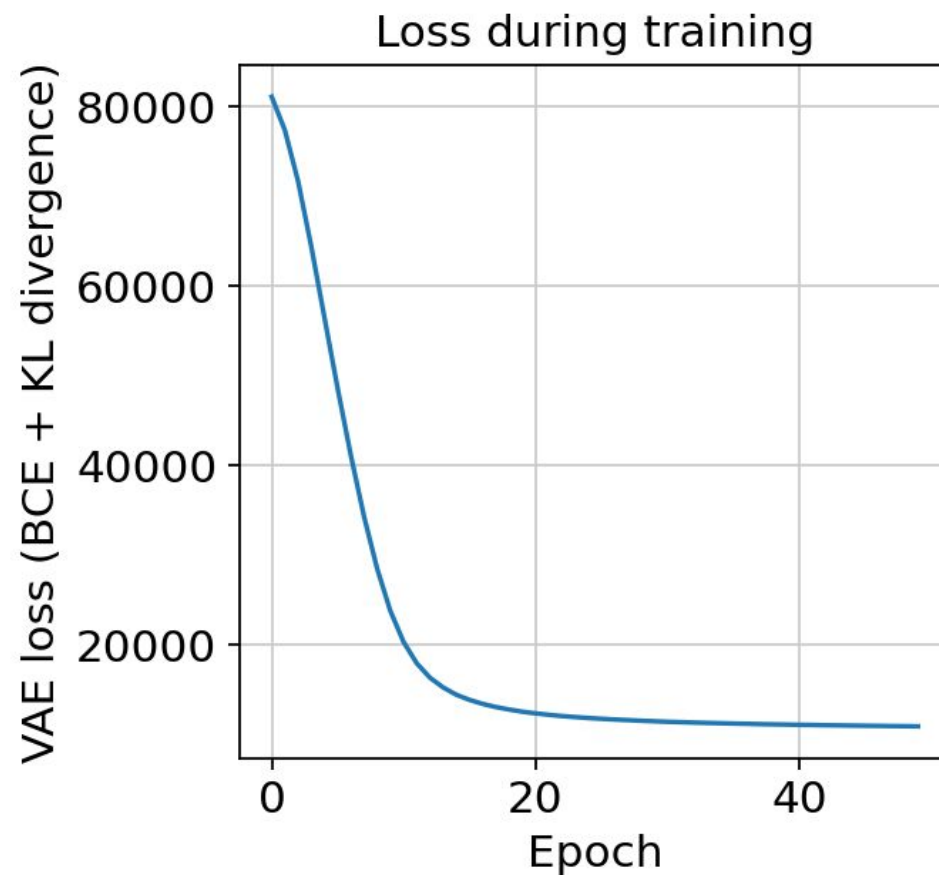
Variational autoencoder



```
1 # Variational Autoencoder model for gene embedding
2 class VAE(nn.Module):
3     def __init__(self, input_size, embedding_size, latent_size):
4         super(VAE, self).__init__()
5         self.fc1 = nn.Linear(input_size, 512)
6         self.fc_mean = nn.Linear(512, latent_size)
7         self.fc_logvar = nn.Linear(512, latent_size)
8         self.fc2 = nn.Linear(latent_size, embedding_size)
9         self.fc3 = nn.Linear(embedding_size, input_size)
10
11     def encode(self, x):
12         x = torch.relu(self.fc1(x))
13         mean = self.fc_mean(x)
14         logvar = self.fc_logvar(x)
15         return mean, logvar
16
17     def reparameterize(self, mean, logvar):
18         std = torch.exp(0.5 * logvar)
19         eps = torch.randn_like(std)
20         return mean + eps * std
21
22     def decode(self, z):
23         z = torch.relu(self.fc2(z))
24         x_hat = torch.sigmoid(self.fc3(z))
25         return x_hat
26
27     def forward(self, x):
28         mean, logvar = self.encode(x)
29         z = self.reparameterize(mean, logvar)
30         x_hat = self.decode(z)
31         return x_hat, mean, logvar
32
```

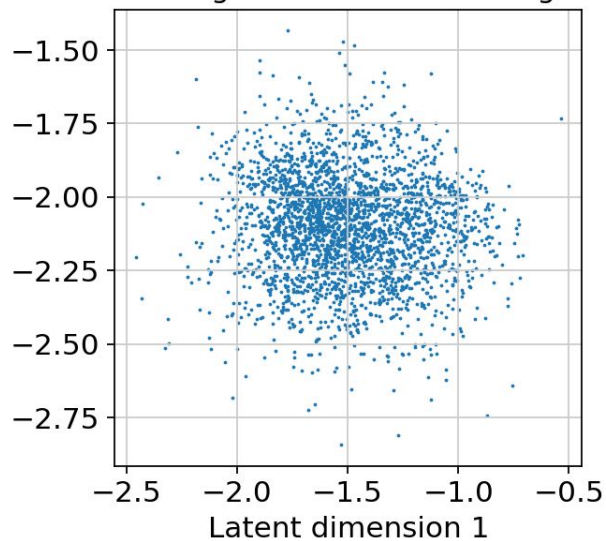
<https://lilianweng.github.io/posts/2018-08-12-vae/>

```
# Define hyperparameters
input_size = count_matrix.shape[1] # Number of genes in the count matrix
embedding_size = 32 # Embedding dimension
latent_size = 2 # Latent variable dimension
learning_rate = 0.001
num_epochs = 50
batch_size = 64
```

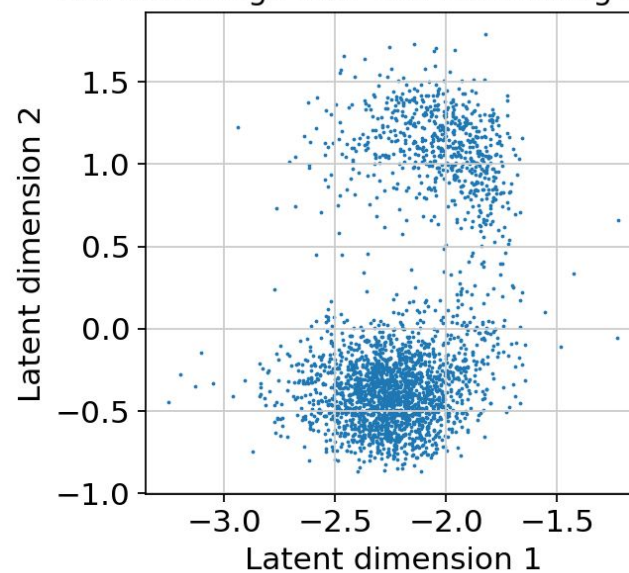


Testing hyperparameter effects on small dataset - 3000 cells

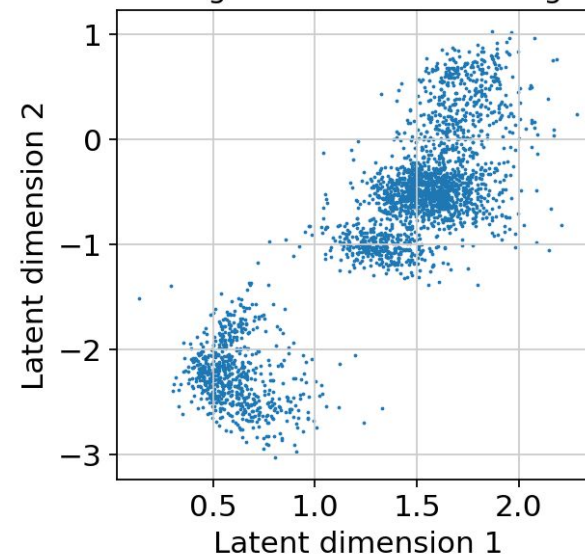
Cell encodings with VAE embedding size=4



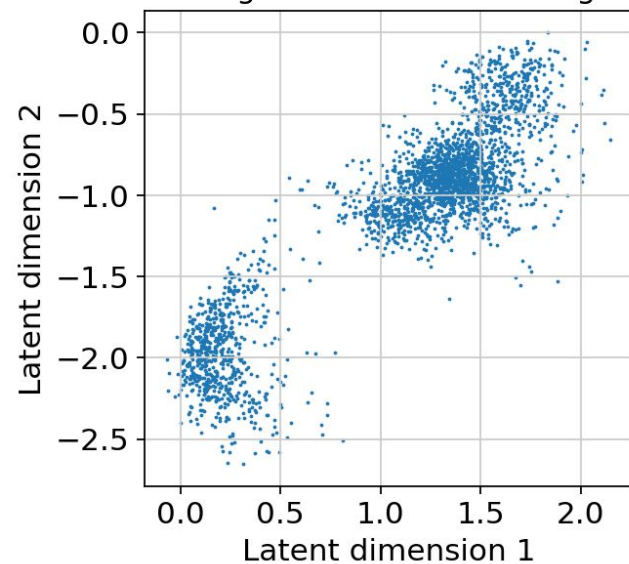
Cell encodings with VAE embedding size=8



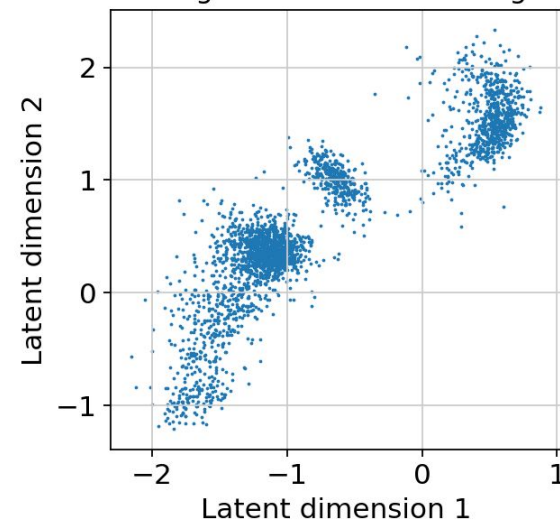
Cell encodings with VAE embedding size=16



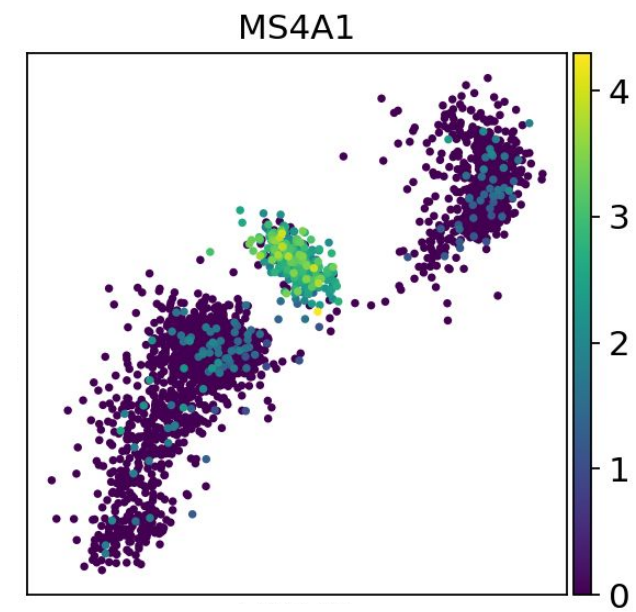
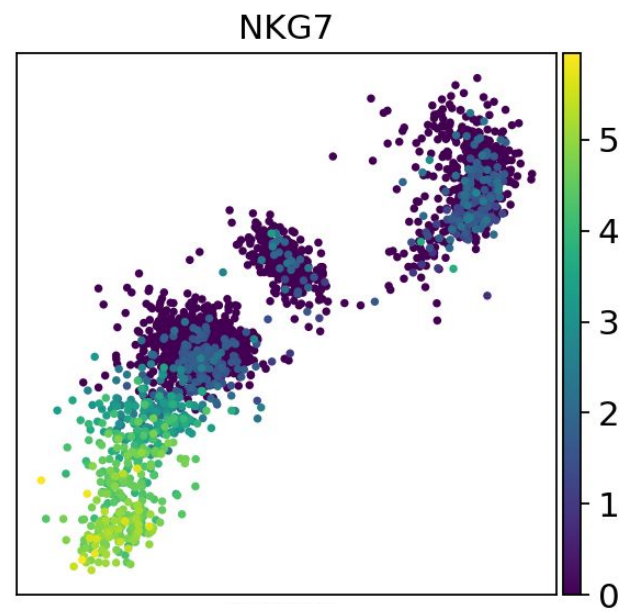
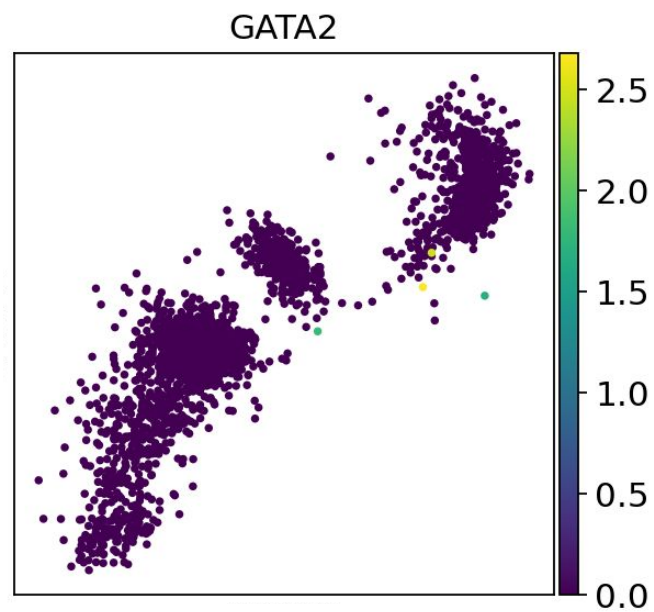
Cell encodings with VAE embedding size=32



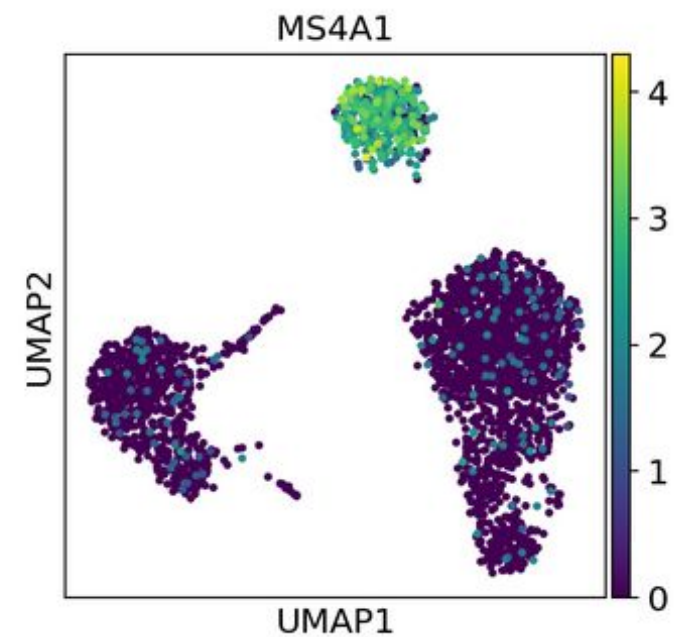
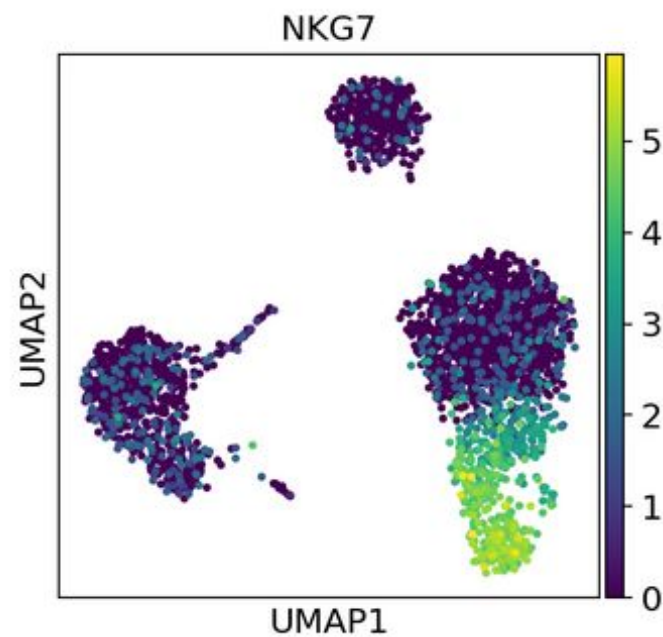
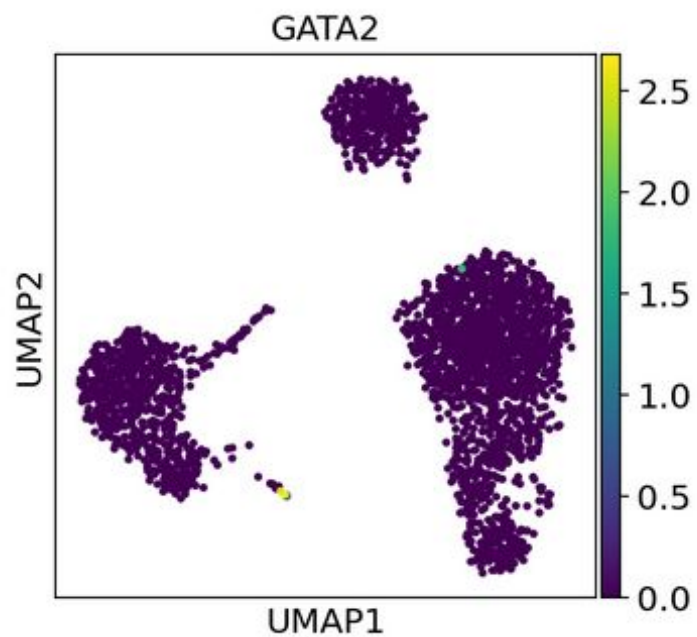
Cell encodings with VAE embedding size=64



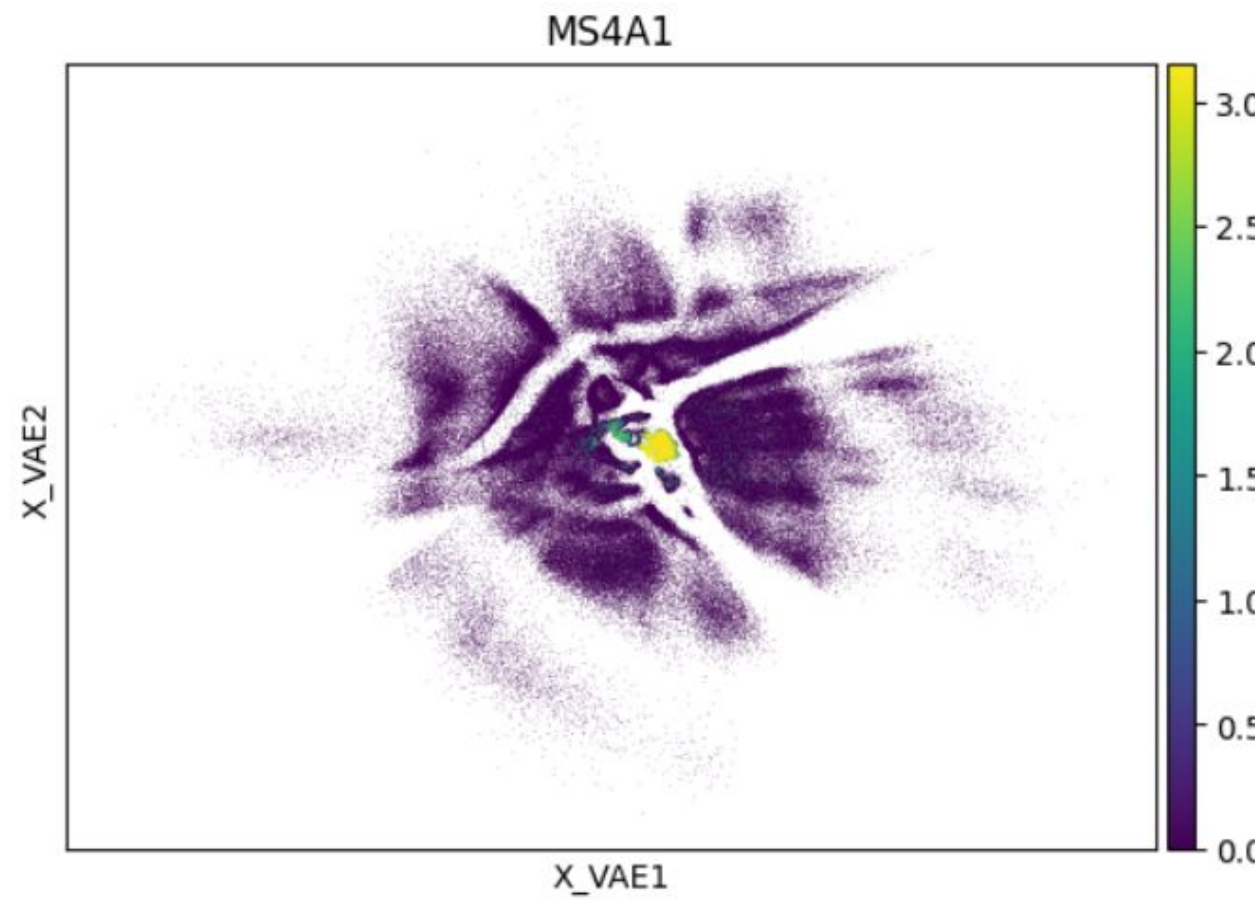
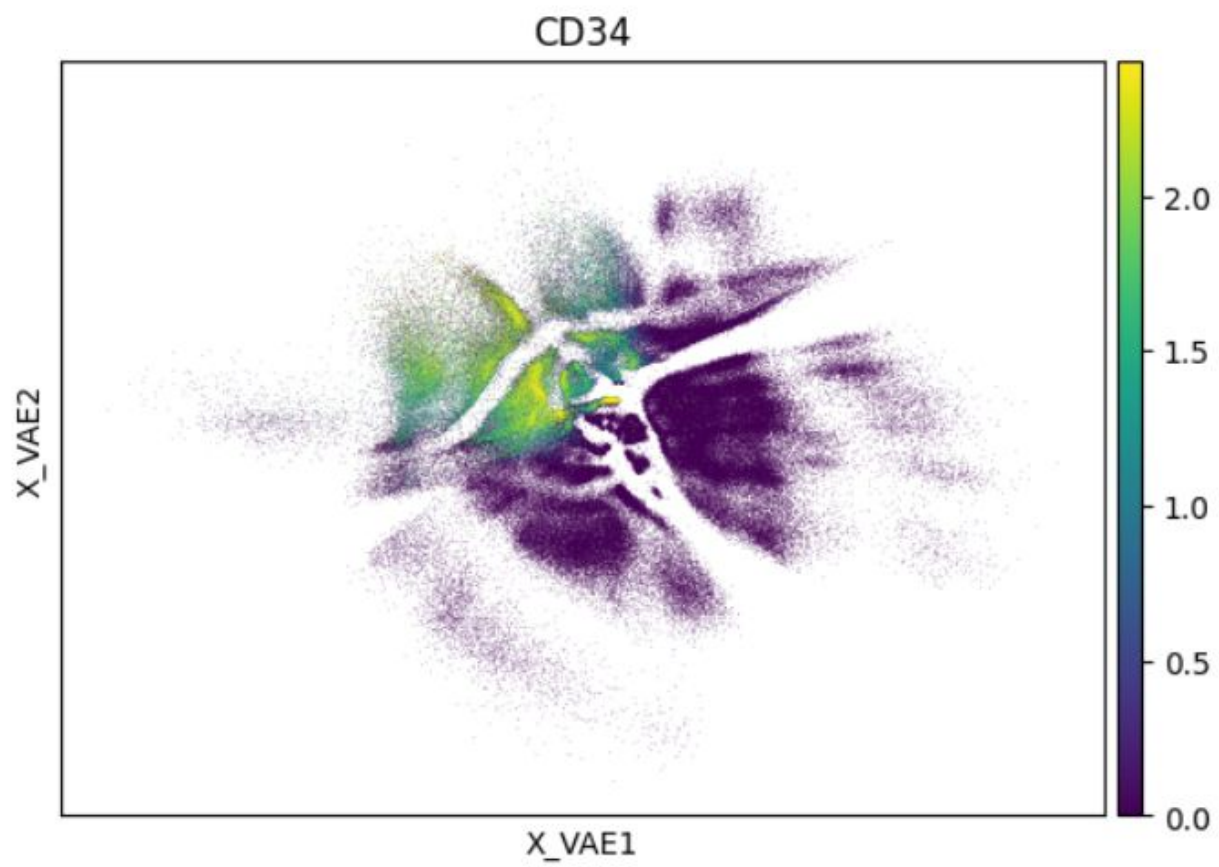
VAE,
embedding
size = 64
latent space



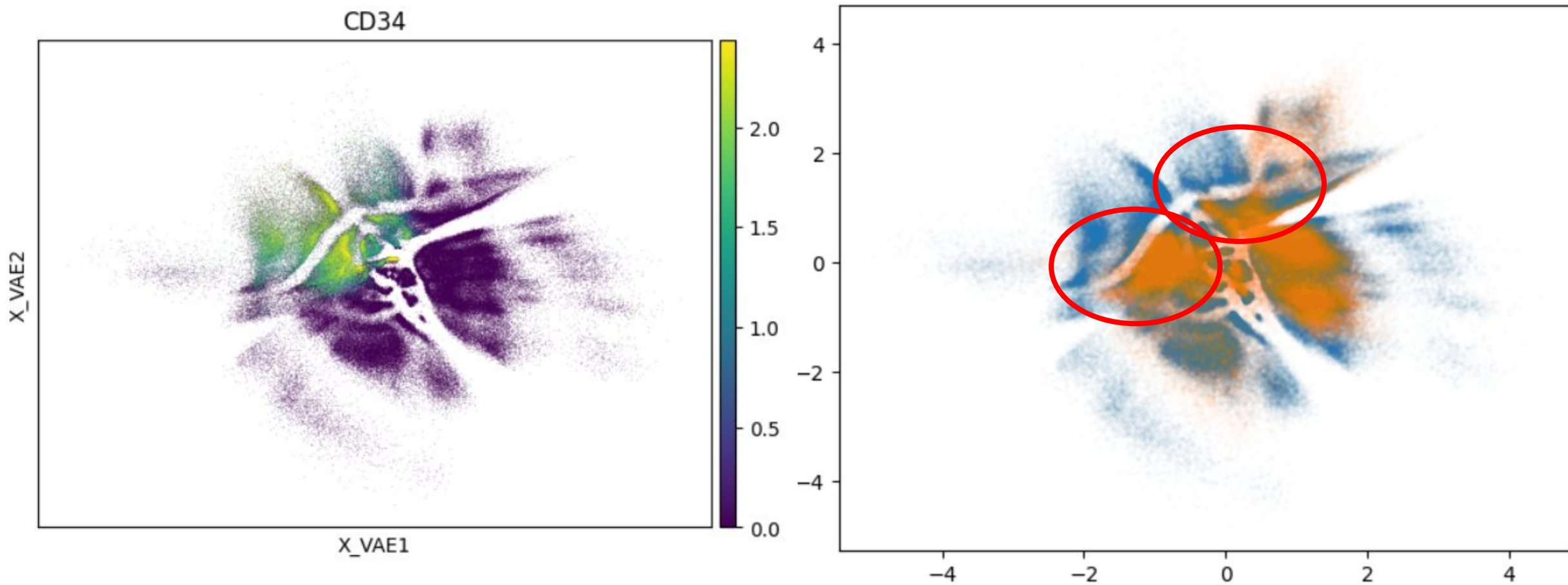
UMAP



Training VAE on large dataset: 263,159 cells



Projecting 152,053 tumor cells on the trained VAE



2 publikacijos su DMSTI afiliacijomis (doktorantūros tezei nebus naudojamos)

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Comprehensive reanalysis for CNVs in ES data from unsolved rare disease cases results in new diagnoses

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